[©]MPI J. ent. Res., 45 (Suppl.) : 1103-1106 (2021)

Potential of the ornamental fish, *Trichopodus trichopterus* (Pallus, 1770) in controlling *Culex* larvae under laboratory condition

Priti Ranjan Pahari^{*}, Gourisankar Mandal, Subhadeep Maiti and Tanmay Bhattacharya^{**}

PG Department of Zoology, Tamralipta Mahavidyalaya, Tamluk - 721 636, West Bengal, India

ABSTRACT

The three spot gourami, *Trichopodus trichopterus*, an ornamental fish, consumed significantly more larvae of *Culex quinquefasciatus* than *Culex quinquefasciatus* pupae and *Chironomus ramosus* larvae irrespective of whether those were offered as prey separately or together in paired combination. This fish predates significantly more number of prey during day time and has significantly higher dietary preference index for *Culex quinquefasciatus* larvae as compared to *Culex quinquefasciatus* pupae or *Chironomus ramosus* larvae. It appears to a very efficient biocontrol agent at least under laboratory condition.

Key words : Biocontrol, *Chironomus ramosus* larvae, *Culex quinquefasciatus* larvae and pupae, dietary preference, larvivorous fish, *Trichopodus trichopterus*.

INTRODUCTION

Fish are more effective controlling agents of the mosquito larvae as compared to copepod predators (Russell et al., 2001) and unlike chemical larvicides mosquito larvae cannot make themselves physiologically resistant to fish (Rodríguez-Pérez et al., 2012). Larvivorous fish are being used in control of mosquito from pre-DDT era (Walker and Lynch, 2007). Raghabendra et al. (2011) have enlisted common mosquito feeding fishes of India. In recent years several papers have been published on the mosquito larvae feeding ability of fishes in India (Phukon and Biswas, 2011, 2013; Bano and Serajuddin, 2016; Das et al., 2018; Pahari et al., 2020 a & b) and elsewhere (Cavalcanti et al., 2007; Seng et al., 2008; Rodríguez-Pérez et al., 2012; Oo et al., 2018). Three spotted gourami, Trichopodus trichopterus (Pallus, 1770) is one of the hardiest fish which can adapt in different water conditions (Sandford, 1999) and in nature it inhabits lowlands, wetlands, marshes and swamps (Kottelat, 2001). Cavalcanti et al. (2007) experimentally demonstrated that it is an efficient predator of Aedes aegypti larvae. In the present paper Culex quinquefasciatus larvae and pupae feeding ability of this fish has been investigated under laboratory condition to evaluate its biocontrol potentiality.

MATERIALS AND METHODS

T. trichopterus specimen were collected from a ornamental fish breeding centre of Tamluk. These were gently placed in a glass aguarium containing filtered pond water and acclimatized for a fortnight before the commencement of experiments. Mosquito larvae and pupae were collected from the drainage system of Tamluk Municipality region. The larvae and pupae were captured by a hand net (mesh size 200 µm). These were transported to the laboratory and kept in an aquarium filled with drain water. Culex quinquefasciatus Say, 1823 larvae and pupae were identified following Tyagi et al. (2015) sorted out and stocked in another aquarium for use in experiments. Chironomus ramosus Choudhuri et al., 1992 were also collected from the drainage system of Tamluk municipality region along with the sediments using trays and baskets, transported and stocked in laboratory.

Two glass aquaria $(30 \times 20 \times 24 \text{ cm})$ were filled each with 6 lit of pond water after passing through a plankton net (mesh size 62 µm) the day before every experiment. Acclimatized fish of approximately similar weight (9.37 - 9.63 g) and length (9.32 - 9.91 cm) were placed one in each experimental aquarium and starved for 24 h before commencement of experiment which lasted for 24 h from 6:00 h in the morning to 6:00 h next day.

^{*}Corresponding author's E-mail : priti.pahari@rediffmail.com **Department of Zoology, Vidyasagar University, Midnapore - 721 102, West Bengal, India

Predation data were recorded at 6 pm (for day time) and 6 am (for night).

Predation efficiency and prey preference were studied by offering prey separately and in paired combination. In the first series in the first set, only Culex guinguefasciatus larvae were given as prey in two aguaria each with one fish. In the second set, only *Culex quinquefasciatus* pupae were given as prey in two aquaria each with one fish. In the third set, only Chironomus ramosus larvae were given as prey in two aquaria each with one fish. Experiment was repeated for four times. In the second series Culex quinquefasciatus larvae and Chironomus ramosus larvae were given together as prey in 1:1 ratio in two aquaria each with one fish. Here also experiments were repeated for four times. Data collected were analysed by using MS-Excel 2013 and IBM SPSS version 25 software. Dietary preference index was computed using the formula of Chesson (1978).

$$\hat{\alpha}_{i} = \frac{\hat{r}_{i}}{\hat{n}_{i}} \left[\frac{1}{\Sigma \left(\frac{\hat{r}_{j}}{\hat{n}_{j}} \right)} \right]$$

[Where, α_i = Manly's alpha (preference index) for prey type i; r_i , r_j = Proportion of prey type i or j in the diet (i and j = 1, 2, 3.... m); n_i , n_j = proportion of prey type i or j in the environment; m = number of prey types possible]

RESULTS AND DISCUSSION

When *T. trichopterus* was offered prey separately it always consumed significantly more (p < 0.001) *Culex quinquefasciatus* larvae as compared to other prey in course of 24 h (Table 1).

The same trend of predation was also observed for day time and night time predation. It was also seen that diurnal consumption was significantly more than the nocturnal predation irrespective of prey type (Table 2).

When prey types were offered together in paired combinations the fish took significantly more *Culex quinquefasciatus* larvae as compared to *Chironomus ramosus* larvae in course of 24 h as well as during day or night (Table 3).

This finding is further supported by the significantly higher dietary preference index of the fish in favour of *Culex quinquefasciatus* larvae (Table 4).

Mosquito larvae feeding ability of various fish species have earlier been investigated by various authors like Cavalcanti *et al.* (2007); Phukon and Biswas (2011, 2013); Bano and Sirajuddin (2016); Oo *et al.* (2018); Pahari *et al.* (2020a & b). Present study revealed that *T. trichopterus* is a predominantly diurnal predator consuming significantly more

Table 1. Predation rate in 24 hrs. of T. trichopterus when prey types were offered separately.

Prey type	No. consumed [Mean ± SE (Range)]	Comparison	t value (p < 0.001)
Culex larvae (ML)	819.88 ± 6.91 (793-852)	ML vs MP	6.32
Culex pupae (MP)	776.63 ± 6.17 (753-802)	ML vs CL	19.92
Chironomus larvae (CL)	647.75 ± 7.17 (628-677)	MP vs CL	24.24

Table 2. Predation rate during day (12 h) and night (12 h) of <i>T. trichopterus</i> when prey types were offered s
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Prey type	Number consumed [Mean ± SE (Range)]		t value (p < 0.001)
	Day (6 am - 6 pm)	Night (6 pm - 6 am)	_
Culex larvae (ML)	472.88 ± 4.31 (459-494)	408.88 ± 4.02 (391-424)	10.85
Culex pupae (MP)	457.13 ± 4.45 (441-476)	389.25 ± 4.73 (373-412)	10.45
Chironomus larvae (CL)	430.38 ± 3.91 (417-446)	368.75 ± 4.13 (353-387)	10.83

Table 3. Predation rate during d	ay and night of <i>T. tr</i>	<i>ichopterus</i> when prey types	were offered in paired combination.

Predation time	Culex quinquefasciatus larvae	Chironomus ramosus larvae	t value
	consumed [Mean ± SE (Range)]	consumed [Mean ± SE (Range)]	(p < 0.001)
Day (6 am - 6 pm)	253.13 ± 3.80 (241-272)	208.50 ± 3.99 (192-227)	8.10
Night (6 pm - 6 am)	227.38 ± 3.84 (212-246)	192.75 ± 3.28 (179-204)	6.85
24 h	490.88 ± 6.08 (471-518)	396.63 ± 5.54 (378-421)	11.46

Ornamental fish, Trichopodus trichopterus in controlling Culex larvae

Predation time	Culex quinquefasciatus larvae	Chironomus ramosus larvae	t value (p < 0.001)
Day (6 am - 6 pm)	0.55	0.45	13.68
Night (6 pm - 6 am)	0.54	0.46	10.19
24 h	0.55	0.45	12.16

Table 4. Comparison of the dietary preference index of T. trichopterus.

prey during day as compared to night. Similar observations have also been made by Phukon and Biswas (2011) for *Chana gachua* and Pahari *et al.* (2020b) for *Stigmatogobius sadanandio*. It appears that *T. Trichopterus* relies more on the visual cues for predation.

Presence and absence of alternative prey adversely affect the mosquito larvae predation ability of a fish. Predatory fish generally seek large and agile prey (Knight et al., 2004; Manna et al., 2008). Devi and Juhari (2011), Barik et al. (2018) and Pahari et al. (2020a & b) have demonstrated the predatory fish have a definite preference for Chironomous larvae over Culex quinquefasciatus larvae. Unlike mosquito larvae, which live mostly at surface in stagnant water, chironomid larvae live at the bottom or on submerged plants and objects (Bay, 2003). Pahari et al. (2020a, 2021) have experimentally demonstrated that the predation ability of Puntius sophore and Colisa fasciata, changes from Chironomous larvae to Culex if the nature of the substratum is made more complex. In the present investigation it has been found that T. Trichopterus always preferred Culex guinguefasciatus larvae than other types of prey. However, there was a relative decline in the predation rate in presence of alternate prey. Similarly presence of additional predators also adversely affects the biocontrol efficiency of a predatory fish (Saha et al., 2010; Pahari et al., 2018).

It may thus be concluded that as *T. Trichopterus* is an efficient predator of *Culex quinquefasciatus* larvae, even in the presence of alternate prey, it may be effectively used in mosquito biocontrol management programme.

ACKNOWLEDGEMENTS

The authors are grateful to the Department of Science & Technology and Biotechnology, Govt. of West Bengal for the financial assistance (Research Project Memo No. 172 (Sanc.)/ST/P/ S&T/1G-70/2017 Dated 16.3.2018). Authors are thankful to the Principal, Tamralipta Mahavidyalaya, Tamluk, Purba Medinipur, West Bengal, India for providing facilities.

REFERENCES

- Bano, F. and Serajuddin, M. 2016. Comparative study of larvicidal efficiency of four indigenous fish with an exotic top water minnow, *Gambusia affinis. J. Ecophysiol. Occup. Hlth.*, **16**: 7-12.
- Barik, M., Bhattacharjee, I., Ghosh, A. and Chandra, G. 2018. Larvivorous potentiality of *Puntius tetrazona* and *Hyphessobrycon rosaceus* against *Culex vishnui subgroup* in laboratory and field based bioassay. *BMC Res. Notes*, **11**: 804, 5 p. DOI: 10.1186/s13104-018-3902-8.
- Bay, E.C. 2003. Chironomid Midges. Washington State University, WSU PLS 45.
- Cavalcanti, L.P. de G., Pontes, R.J.S., Regazzi, A.C.F., Júnior F.J. de P., Frutuoso, R.L., Sousa, E.P., Filho, F.F.D. and Lima, J.W. de O. 2007.
 Efficacy of fish as predators of *Aedes aegypti* larvae, under laboratory conditions. *Rev. Saúde Pública*, **41**: 638-44. DOI: 10.1590/S0034-89102006005000041.
- Das, M.K., Rao, M.R.K. and Kulsreshtha, A.K. 2018. Native larvivorous fish diversity as a biological control agent against mosquito larvae in an endemic malarious region of Ranchi district in Jharkhand, India. *J. Vector Borne Dis.*, **55**: 34-41.
- Devi, N.P. and Jauhari, R.K. 2011. Food preference of *Aplocheilus panchax* (Cyprinidontiformes: Aplocheilidae) with special reference towards mosquito larvae. *Researcher*, **3**: 55-59. DOI: 10.7537/marsrsj030611.10.
- Knight, T.M., Chase, J.M., Goss, C.W. and Knight, J.J. 2004. Effects of interspecific competition, predation and their interaction on survival and development time of immature

Anopheles quadrimaculatus. J. Vector Ecol., **29**: 277-84.

- Kottelat, M. 2001. Fishes of Laos. WHT Publications Ltd., Colombo 5, 198 p.
- Manna, B., Aditya, G. and Banerjee, S. 2008. Vulnerability of the mosquito larvae to the guppies (*Poecilia reticulata*) in the presence of alternative preys. *J. Vector Borne Dis.*, **45**: 200-206.
- Oo, N.N., Thone, M.T., Ko, M.M.M. and Mya, M.M. 2018. Biological control of *Aedes* larvae using indigenous fish *Rasbora daniconius* (Nga Dawn Zin) and *Colisa fasciata* (Nga Thit Kyauk) from Pakokku Township, Magwe Region. *J. Biol. Engg. Res. Rev.*, **5**: 01-08.
- Pahari, P.R., Chakrabortty, D., Mandal, B. and Bhattacharya, T. 2018. Biological control of mosquito larvae using naiad of Ruddy Marsh Skimmer *Crocothemis servilia*. Indian J. Ent., 80: 1503-505. DOI: 10.5958/0974-8172.2018.00330.9
- Pahari, P.R., Maiti, S., Mandal, S.S., Bhattacharya, R.P. and Bhattacharya, T. 2020a. *Culex quinquefasciatus* Say, 1823 larvae feeding ability of *Puntius sophore* (Hamilton, 1822) in laboratory condition. *J. ent. Res.*, **44**: 103-106. DOI: 10.5958/0974-4576.2020.00019.5
- Pahari, P.R., Mishra, N.P., Sahoo, A., Bhattacharya, R.P. and Bhattacharya, T. 2020b. First record of the mosquito control potentiality of *Stigmatogobius sadanundio* (F. Hamilton, 1822) Gobiidae, Perciformes in laboratory condition. *Trop Parasitol.*, **10**: 130-35. DOI: 10.4103/ tp.TP_55_19.
- Pahari, P.R., Mandal, S.S., Maiti, S., Mandal, S. and Bhattacharya, T. 2021. A study on the *Culex quinquefasciatus* Say, 1823 control potentiality of *Colisa fasciata* (Bloch & Schneider, 1801) in Laboratory Condition. *Indian J. Public Health Res. Develop.*, **12**: 144-49.

- Phukon, H.K. and Biswas, S.P. 2011. Investigation on *Channa gachua* as a potential biological control agent of mosquitoes under laboratory conditions. *Asian J. Exp. Biol. Sci.*, **2**: 606-11.
- Phukon, H.K. and Biswas, S.P. 2013. An investigation on larvicidal efficacy of some indigenous fish species of Assam, India. *Adv. Biores.*, **4**: 22-25.
- Raghabendra, K., Barik, T.K., Reddy, B.P.N., Sharma, P. and Dash, A.P. 2011. Malaria vector control: from past to future. *Parasitol. Res.*, **108**: 757-79. DOI: 10.1007/s00436-010-2232-0.
- Rodríguez-Pérez, M.A., Howard, A.F.V. and Reyes-Villanueva, F. 2012. Biological control of dengue vectors. Integrated pest management and pest control – current and future tactics, pp. 241-270. DOI: 10.5772/31589
- Russell, B.M., Wang, J., Williams, Y., Hearnden, M.N. and Kay, B.H. 2001. Laboratory evaluation of two native fishes from tropical north queensland as biological control agents of subterranean Aedes aegypti. J. American Mosquito Contrl. Assoc., 17: 124-26.
- Saha, N., Aditya, G., Saha, G.K. and Hampton, S.E. 2010. Opportunistic foraging by heteropteran mosquito predators. *Aquat. Ecol.*, **44**: 167-76. DOI: 10.1007/s10452-009-9250-y
- Sandford, G. 1999. Aquarium : owner's guide. New York: DK Pub. 264 p.
- Seng, C.M., Setha, T., Nealon, J., Socheat, D., Chantha, N. and Nathan, M.B. 2008. Communitybased use of the larvivorous fish *Poecilia reticulata* to control the dengue vector *Aedes aegypti* in domestic water storage containers in rural Cambodia. J. Vector Ecol., **33**: 139-44.
- Walker, K. and Lynch, M. 2007. Contributions of Anopheles larval control to malaria suppression in tropical Africa: review on achievements and potential. *Med. Vet. Ent.*, **21**: 2-21.

(Received : July 16, 2021; Accepted : December 12, 2021)